

Project 1: *Molecular Genetics Approaches to Developing Scab Resistant Barley.*

1. What are the major goals and objectives of the project?

Previous work in my laboratory has resulted in identifying a barley UDP-glucosyltransferase (HvUGT13248) that exhibits resistance to FHB and trichothecenes when expressed in transgenic wheat, and fine mapping a QTL for FHB resistance in barley on chromosome 6H bin 7. The major goals of this grant were to develop germplasm resources and tools to increase FHB resistance in barley. The specific objectives are (1) to characterize transgenic barley overexpressing *HvUGT13248*; and (2) fine map and characterize the chromosome 6H bin 7 FHB resistance QTL.

2. What was accomplished under these goals? *Address items 1-4) below for each goal or objective.*

1) major activities

Objective 1. Characterize transgenic barley overexpressing HvUGT13248. We created transgenic barley lines overexpressing *HvUGT13248* and showed that they exhibit high levels of DON resistance in roots on DON-containing media. To generate materials that can be screened in the field, we backcrossed the *HvUGT13248* transgene into Rasmusson and selected lines that are homozygous for the transgene. Transgenic plants carrying the *HvUGT13248* transgene in the Rasmusson background were planted in the field in the summer of 2018 and inoculations and scoring are ongoing. The field trials are a collaboration with Ruth Dill-Macky.

Objective 2. Fine map and characterize the chromosome 6H bin7 FHB resistance QTL. Barley QTL associated with Fusarium head blight resistance, reduced deoxynivalenol accumulation and increased grain protein colocalize on the short arm of chromosome 6H bin 7. To understand the complex genetics of this QTL, we are conducting a fine mapping project. We generated a large F₂ segregating population (~2,000 individuals) from crossing a near-isogenic line carrying the chromosome 6H bin 7 resistant allele in the cultivar Lacey genetic background to Lacey. SSR markers were used to identify recombinants in the chromosome 6H bin 7 region from the F₂ population, which were further genotyped with 34 SNP markers to identify 13 recombinant classes. Homozygous recombinants in the F_{2:3} families were identified with SNP markers and homozygous F₄ plants were tested in field trials in St. Paul in 2016 and 2017 for FHB severity, DON accumulation and grain protein content. All data (FHB severity, DON accumulation, and grain protein content) have been collected from the trials. From the 2016 field test, we identified recombinants that exhibit resistance that appears to be uncoupled from high grain protein content. In 2017, the disease pressure was too low to obtain reliable FHB data. The same lines have been planted in the field in 2018 and inoculations and disease scoring are ongoing.

Three additional activities related to this project include: A bi-parental F_{6:7} RIL population was developed from Rasmusson crossed to PI383933 and used to map QTL for FHB resistance, reduced DON accumulation and other agronomic traits. PI383933 is a highly susceptible landrace that exhibits early heading date, short stature and dense spikes. The population was phenotyped in St. Paul, MN and Crookston, MN in 2015 and in 2016 and genotyped with the iSELECT 9K barley chip. QTL analysis identified six QTL for FHB severity and DON accumulation on chromosomes 2H, 3H, 5H, 6H and 7H with the largest effect QTL located on chromosome 7H. Three of the QTL on chromosome 3H, 5H and 6H appear to be novel. A manuscript describing these results was submitted to *Frontiers in Plant Science*.

We analyzed RNA-Seq data from two NILs carrying FHB resistant alleles at chromosome 2H bin8 and chromosome 6H bin7, and their recurrent parents. We identified differentially expressed genes between the plants carrying the resistant and susceptible alleles. These genes are helping us understand and manipulate resistance in barley. We published the results of this study in *BMC Genomics* (Huang et al., 2016) using previous funding from the USWBSI. The RNA-seq data were also used to identify SNPs between the NILs and recurrent parents for the fine mapping efforts of the chromosome 2H bin8 and 6H bin7 QTL, and for a fine mapping effort in Kevin Smith's laboratory.

A major barley FHB QTL is located in the chromosome 2H bin8 region. To fine map this region, an F₂ population was generated from near-isogenic lines in the M69 genetic background carrying the resistant allele crossed to M69, a susceptible line. Two KASP SNP markers were used to genotype ~2,000 plants to identify recombinants. To determine the general location of the breakpoints, the recombinants were genotyped with another 33 SNP markers within the introgressed region. Homozygous F₃ plants were phenotyped for FHB resistance, heading date, and DON accumulation in St. Paul in 2016 and 2017. Unfortunately, the disease pressure was low in 2017 and the data were unreliable. Lines that exhibit reduced FHB severity and early heading date were identified in 2016 and are currently being retested.

2) specific objectives

Objective 1. Characterize transgenic barley overexpressing *HvUGT13248*.

Objective 2. Fine map and characterize the chromosome 6H bin 7 FHB resistance QTL.

3) significant results

We have identified recombinants in the chromosome 6H bin7 and chromosome 2H bin8 regions and are in the process of fine mapping each region. We developed transgenic barley overexpressing *HvUGT13248* that exhibits resistance in roots in DON-containing media. Backcross lines in the Rasmusson background containing the *HvUGT13248* transgene have been developed. We mapped three novel QTL for FHB resistance on chromosome 3H, 5H and 6H.

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4) Key outcomes or other achievements

Our preliminary fine mapping results indicate that we have recombinants that contain FHB resistance without the deleterious high grain protein content and late heading date alleles. We developed transgenic barley overexpressing *HvUGT13248* that exhibits resistance to DON and introgressed the *HvUGT13248* transgene into Rasmusson. Also, three novel QTL were detected that are associated with resistance to FHB.

3. What opportunities for training and professional development has the project provided?

A Ph.D. student and a Postdoctoral Research Associate have worked on this project. The Ph.D. student graduated in summer of 2017 and is now a postdoctoral research associate at North Carolina State University. Both have presented their work at the National Scab Forum and the postdoc presented his work at the International Barley Genetics Symposium. The postdoc and graduate student meet with me regularly and participate in weekly lab meetings.

4. How have the results been disseminated to communities of interest?

We presented our work at a poster at the National Scab Forum. A paper has been submitted to *Frontiers in Plant Science* describing mapping FHB resistance in the Rasmusson x PI383933 population.

Project 2: *Characterizing Trichothecene Resistance and Developing Scab Resistant Wheat.*

1. What are the major goals and objectives of the project?

Fusarium head blight (FHB, scab), a fungal disease of small grain crops caused by *Fusarium graminearum*, threatens to reduce wheat and barley to economically unviable crops in the United States. During infection the fungus produces trichothecene mycotoxins such as deoxynivalenol (DON) that have been shown to increase fungal virulence. To complement the current breeding efforts, a major goal of my laboratory is to develop and characterize transgenic wheat exhibiting trichothecene and FHB resistance. Previously, my laboratory developed transgenic wheat carrying a barley UDP-glucosyltransferase (*HvUGT13248*) and showed that these transgenics exhibit high levels of FHB resistance via conjugation of DON to DON-3-O-glucoside (D3G). There are three major objectives in the proposed work including: (1) develop elite wheat cultivars with FHB resistance; (2) characterize the accumulation of trichothecenes and trichothecene-glucoside conjugates in transgenic wheat carrying *HvUGT13248*; and (3) test potential trichothecene resistance genes.

2. What was accomplished under these goals? *Address items 1-4) below for each goal or objective.*

1) Major activities

Objective 1. Develop elite wheat cultivars with FHB resistance. We backcrossed the *HvUGT13248* transgenic line into the cultivar Linkert and identified six homozygous lines with transgene expression, and ten lines without transgene expression. We also developed backcross lines of *HvUGT13248* transgenics in the cv. Rollag genetic background and identified four lines of each of the four genotypes, namely *UGT+/Fhb1+*, *UGT-/Fhb1+*, *UGT+/Fhb1-*, and *UGT-/Fhb1-* from four different transgenic events (a total of 64 lines). These lines were screened in the greenhouse in the Fall 2016 and spring 2017. In the Rollag background, lines carrying the combination of *HvUGT13248* and *Fhb1* exhibited stable and higher resistance than *Fhb1* alone. In the Linkert background, lines carrying *HvUGT13248* exhibit higher resistance than lines that did not carry the transgene. The lines were planted in the field in 2018 and inoculations and scoring is ongoing.

Objective 2. Characterize the accumulation of trichothecenes and trichothecene-glucoside conjugates in transgenic wheat carrying *HvUGT13248*. We showed that transgenic wheat expressing *HvUGT13248* exhibits high levels of resistance to DON-producing *F. graminearum* strains due to the conjugation of DON to DON-3-O-glucoside (Li et al., 2015). We also showed that these same transgenic wheat lines exhibit high levels of type II resistance to NIV-producing *F. graminearum* and the transgenic wheat quickly converts NIV to NIV-3-O-glucoside. A paper was published that described the NIV detoxification results (Li et al., 2017). These lines also exhibit resistance to three other trichothecenes (3,15-di-ANIV, NX-2, and 3-ADON).

Objective 3. Test potential trichothecene resistance genes. To rapidly identify additional DON resistance genes, we transformed *Arabidopsis* with putative DON

resistance genes from barley and tested the transgenics on DON-containing media. We transformed Arabidopsis with a zinc finger protein, two ABC transporters, two cytochrome P450s, one epoxide hydrolase, three glutathione-S-transferases and a cysteine synthase. We did not identify any genes that resulted in increased DON resistance.

2) Specific objectives

Objective 1. Develop elite wheat cultivars with FHB resistance.

Objective 2. Characterize the accumulation of trichothecenes and trichothecene-glucoside conjugates in transgenic wheat carrying *HvUGT13248*.

Objective 3. Test potential trichothecene resistance genes.

3) Significant results

We developed transgenic wheat in elite cultivars that may provide enhanced resistance to FHB and will provide the genetic materials to study the potential interactions between *HvUGT13248* and *Fhb1*. Our greenhouse tests showed that the combination of *HvUGT13248* and *Fhb1* in the same background exhibited stable and higher resistance than *Fhb1* alone. We have also shown that *HvUGT13248* provides resistance to a broad range of trichothecene mycotoxins including: DON, NIV, 3,15-di-ANIV, NX-2, and 3-ADON.

4) key outcomes or achievements

We developed transgenic wheat lines that exhibit resistance to FHB and to a broad spectrum of trichothecenes. The transgene, *HvUGT13248*, has been introgressed into two elite cultivars and we tested those lines in the greenhouse and showed that the combination of *HvUGT13248* and *Fhb1* in the same background exhibited stable and higher resistance than *Fhb1* alone.

3. What opportunities for training and professional development has the project provided?

A Ph.D. student has worked on this project and he graduated in the summer of 2017. Currently, he is a postdoc at North Carolina State University. He has presented his work at the National Scab Forum, participates in weekly lab meetings, and meets regularly with me.

4. How have the results been disseminated to communities of interest?

We published a manuscript in Journal of Experimental Botany (Li et al., 2017) describing some of this work.

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Training of Next Generation Scientists

Instructions: Please answer the following questions as it pertains to the FY17 award period. The term “support” below includes any level of benefit to the student, ranging from full stipend plus tuition to the situation where the student’s stipend was paid from other funds, but who learned how to rate scab in a misted nursery paid for by the USWBSI, and anything in between.

1. **Did any graduate students in your research program supported by funding from your USWBSI grant earn their MS degree during the FY17 award period?** No

If yes, how many?

2. **Did any graduate students in your research program supported by funding from your USWBSI grant earn their Ph.D. degree during the FY17 award period?** Yes

If yes, how many? 1

3. **Have any post docs who worked for you during the FY17 award period and were supported by funding from your USWBSI grant taken faculty positions with universities?** No

If yes, how many?

4. **Have any post docs who worked for you during the FY17 award period and were supported by funding from your USWBSI grant gone on to take positions with private ag-related companies or federal agencies?** No

If yes, how many?

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Release of Germplasm/Cultivars

Instructions: In the table below, list all germplasm and/or cultivars released with full or partial support through the USWBSI during the FY17 award period. All columns must be completed for each listed germplasm/cultivar. Use the key below the table for Grain Class abbreviations. *Leave blank if you have nothing to report or if your grant did NOT include any VDHR-related projects.*

Name of Germplasm/Cultivar	Grain Class	FHB Resistance (S, MS, MR, R, where R represents your most resistant check)	FHB Rating (0-9)	Year Released
None to report				

Add rows if needed.

NOTE: List the associated release notice or publication under the appropriate sub-section in the ‘Publications’ section of the FPR.

Abbreviations for Grain Classes

- Barley - BAR
- Durum - DUR
- Hard Red Winter - HRW
- Hard White Winter - HWW
- Hard Red Spring - HRS
- Soft Red Winter - SRW
- Soft White Winter - SWW

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Publications, Conference Papers, and Presentations

Instructions: Refer to the FY17-FPR_Instructions for detailed instructions for listing publications/presentations about your work that resulted from all of the projects included in the FY17 grant. Only include citations for publications submitted or presentations given during your award period (5/17/17 - 5/16/18). If you did not have any publications or presentations, state 'Nothing to Report' directly above the Journal publications section.

NOTE: Directly below each reference/citation, you must indicate the Status (i.e. published, submitted, etc.) and whether acknowledgement of Federal support was indicated in publication/presentation.

Journal publications.

Li, X., H. Michlmayr, W. Schweiger, A. Malachova, S. Shin, Y. Huang, Y. Dong, G. Wiesenberger, S. McCormick, M. Lemmens, P. Fruhmann, C. Hametner, F. Berthiller, G. Adam and G.J. Muehlbauer. 2017. A barley UDP-glucosyltransferase inactivates nivalenol and provides Fusarium head blight resistance in transgenic wheat. *J. Exp. Bot.* 68:2187-2197.

Status: Published

Acknowledgement of federal support: YES

Huang, Y., M. Haas, S. Heinen, B.J. Steffenson, K.P. Smith, and G.J. Muehlbauer. QTL mapping of Fusarium Head Blight resistance and correlated agromorphological traits in an elite barley cultivar Rasmusson.

Status of publication: Submitted to *Frontiers of Plant Science*, under review

Acknowledgement of federal support: YES

Books or other non-periodical, one-time publications.

None

Other publications, conference papers and presentations.

Huang, Y., S. Heinen, B. Steffenson, K.P. Smith, and G.J. Muehlbauer. 2017. Mapping FHB quantitative trait loci in barley. In: S. Canty, A. Clark, S. Vukasovich and D. Van Sanford (Eds.), *Proceedings of the 2017 National Fusarium Head Blight Forum*. Milwaukee, WI, US.

Status: Abstract published and poster presented

Acknowledgement of Federal Support: YES (poster)