

**Project FY24-PB-001:** Explore RNAi to control FHB and mycotoxin contamination

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**1. What are the major goals and objectives of the research project?**

The goal of this project is to develop an endophytic fungal RNAi delivery platform to reduce FHB and mycotoxin contamination.

The objectives of this proposal are:

- **Objective 1:** Design and generate *F. graminearum* RNAi mutants targeting genes that are essential for its pathogenesis, and trichothecene and zearalenone biosynthesis, and determine their effects on toxin production and FHB severity.
- **Objective 2:** Build, evaluate, and optimize the *S. zeae*-mediated RNAi delivery system. We will generate *S. zeae* GFP-RNAi strains, examine RNAi molecule production and the transferring of RNAi signals from *S. zeae* to plants and *F. graminearum*, and determine gene silencing efficacy.

**2. What was accomplished under these goals or objectives? (For each major goal/objective, address these three items below.)**

**What were the major activities?**

**Objective1**

- We performed wheat heads treatment with *S. zeae* strain with different concentrations and examined disease and mycotoxin reduction efficacy.
- We performed wheat seeds treatment with *S. zeae* TRI5 strain with different concentrations and determined RNAi efficacy.

**Objective 2:**

- In collaboration with Dr. Innes from Indiana University, we isolated dsRNA and sRNA from cultures and media grown *S. zeae* GFP RNAi strain, performed Northern Blot. As a control, we also transformed *Fusarium graminearum* PH-1 strain with GFP RNAi construct, isolated sRNA and conducted Northern blot.

**What were the significant results?**

We confirmed that *S. zeae* GFP RNAi strain can produce and secrete dsRNA. Using PH-1 strain expressing GFP RNAi as a control, we confirmed *S. zeae* can produce dsRNA but could not process dsRNA to sRNA.

**List key outcomes or other achievements.**

While we worked on *S. zeae* RNAi strain. We discovered that *S. zeae* alone can suppress 15-ADON production in cultures. In collaboration with Dr. Susan McCormick, we identified a secondary metabolite that can suppress *Fusarium* growth and toxin production. Further investigations are underway to check its ability to reduce fungal growth and toxin production by seed treatments.

**3. What opportunities for training and professional development has the project provided?**

One ORISE fellow, Nick Rhodes, has been trained in molecular biology, such as RNA and sRNA isolation, generation GFP-labelled fungal strain, confocal microscope to investigate fungal colonization. He was also trained in seed treatment, fungal inoculation and FHB virulence assays.

**4. How have the results been disseminated to communities of interest?**

Poster presentation at the 2024 National Fusarium Head Blight Forum, Dec. 7-11, 2024. Austin, TX

**5. What do you plan to do during the next reporting period to accomplish the goals and objectives?**

Funding for this project was terminated. We will summarize the data and prepare a manuscript for publication. In addition, we will do some follow up studies on the secondary metabolite we identified from *S. zeae* for potential FHB and toxin control.